

Supplementary Materials:

Figure S1. Selective inhibition of ROR γ by VTP-938.

Figure S2. Gating strategy for CD4⁺ T cells in lung-draining lymph nodes.

Figure S3. Identification by CyTOF of cell populations in the lungs of OVA-challenged mice.

Figure S4. Gating strategy for IL-17-producing cells in the lung.

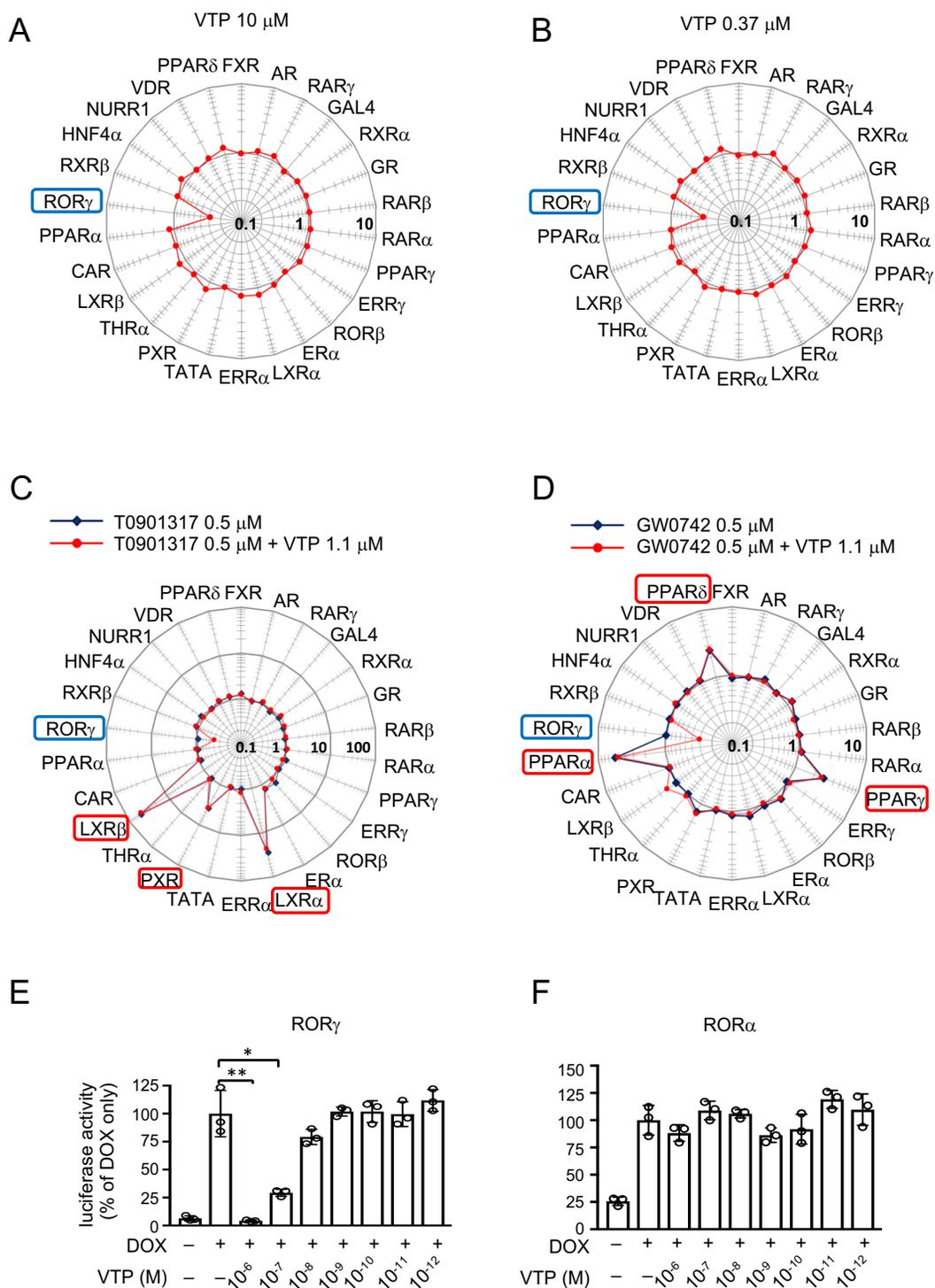
Figure S5. Confocal image of PCLS showing localization of Th17 cells.

Figure S6. Effect of VTP given prior to allergen challenge on three consecutive days.

Figure S7. Identification by CyTOF of cell populations in the lungs of HDE-challenged mice.

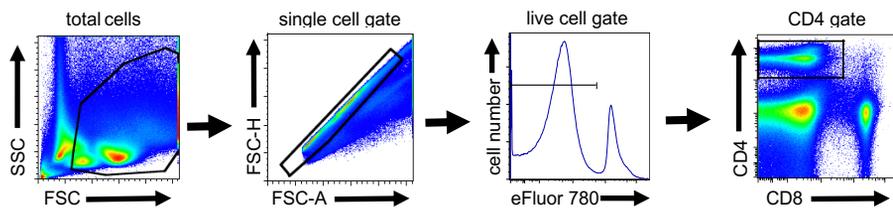
Figure S8. Plasma concentrations of VTP-938.

Table S1. List of antibodies used for CyTOF experiments.



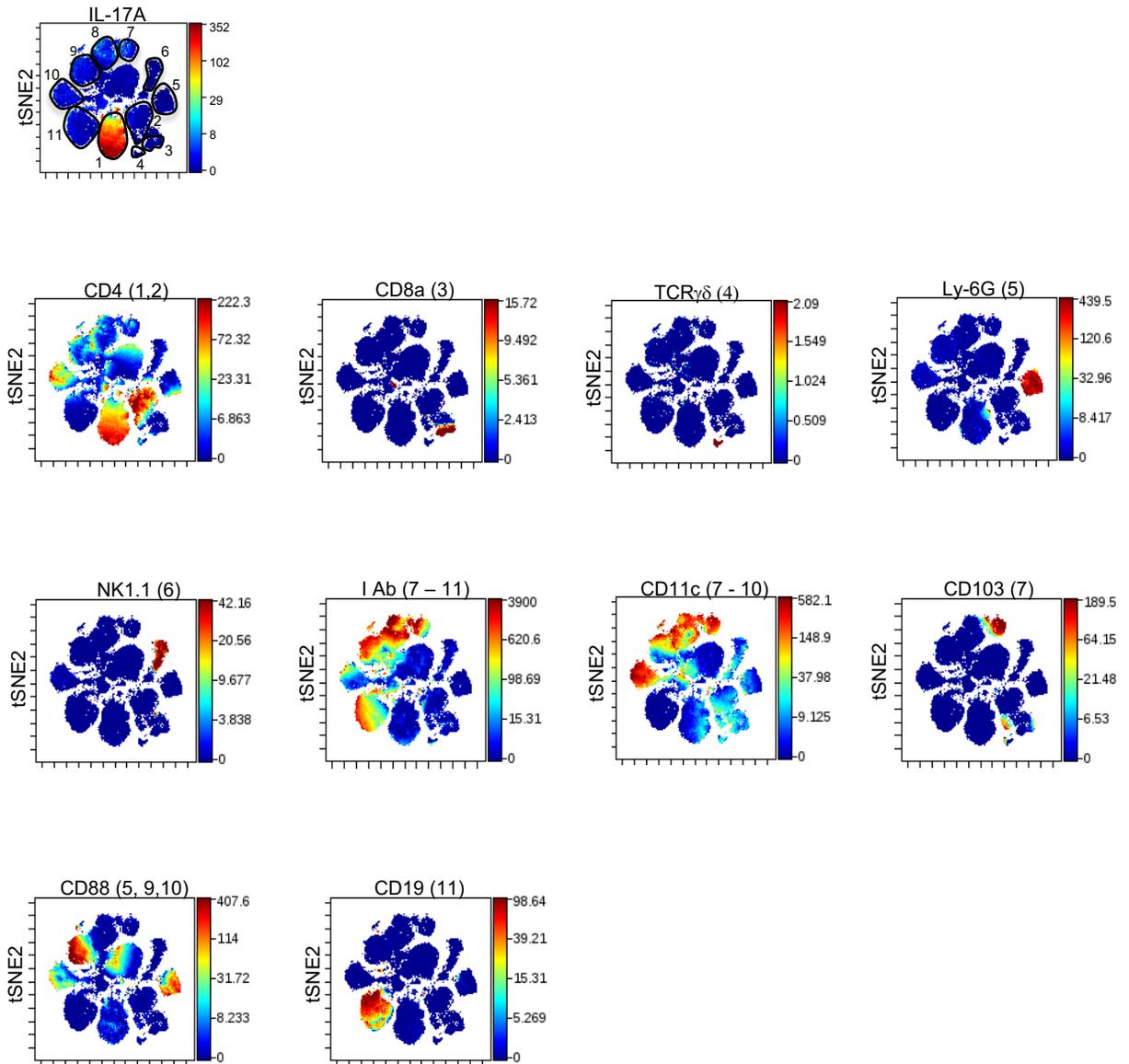
Supplementary Figure S1. Selective inhibition of ROR γ by VTP-938. (A, B) Trans-Factorial assay assessing the effect of VTP (10 or 0.37 μ M) on the activity of 24 human nuclear receptors. VTP only inhibits ROR γ . (C) Trans-Factorial assay examining the effect of VTP (1.1 μ M) on the activation of nuclear receptors by the LXR agonist T0901317. "1", indicates VTP has no effect; "10", 10-fold increase; "0.1", 10-fold decrease; "100", 100-fold increase. VTP has no effect on the activation of LXR α , PXR, and LXR β . (D) Trans-Factorial assay examining the effect of VTP (1.1 μ M) on the activation of nuclear receptors by the PPAR agonist GW0742. VTP has no effect on the activation of PPAR α , PPAR β , and PPAR γ . (E, F) Doxycycline (DOX)- and ROR-dependent activation of a RORE-driven Luciferase (LUC) reporter in CHO ROR γ -Tet-On cells (E) and CHO ROR α -Tet-On cells (F) in the presence of different VTP concentrations. Shown is percent LUC activity relative to cells treated with DOX plus vehicle (DMSO) alone (bar 2). *P<0.05; **P<0.01, as determined by 2-way ANOVA.

Supplementary Figure S2



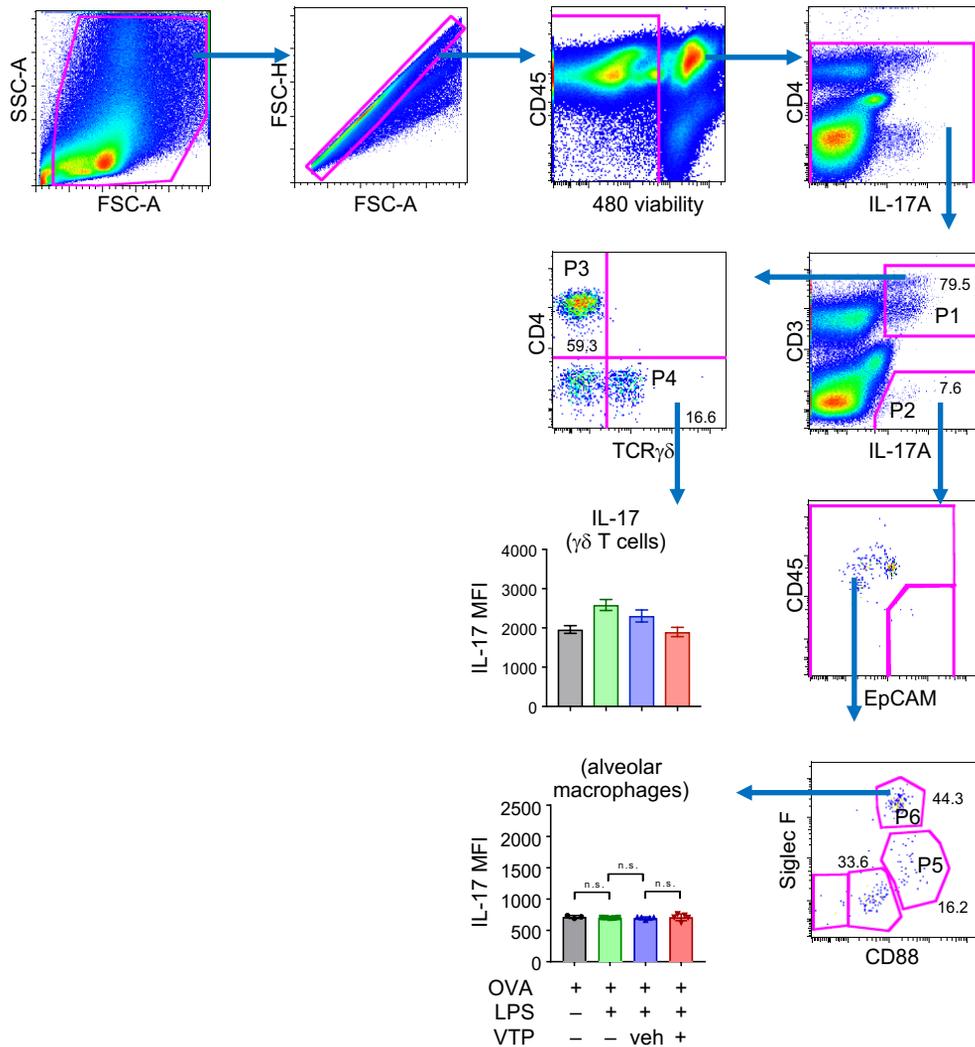
Supplementary Figure S2. Gating strategy for CD4⁺ T cells in lung-draining lymph nodes. Gates are based on forward and side scatter (FSC and SSC), single cells, live cells, and CD4 and CD8 staining.

Supplementary Figure S3



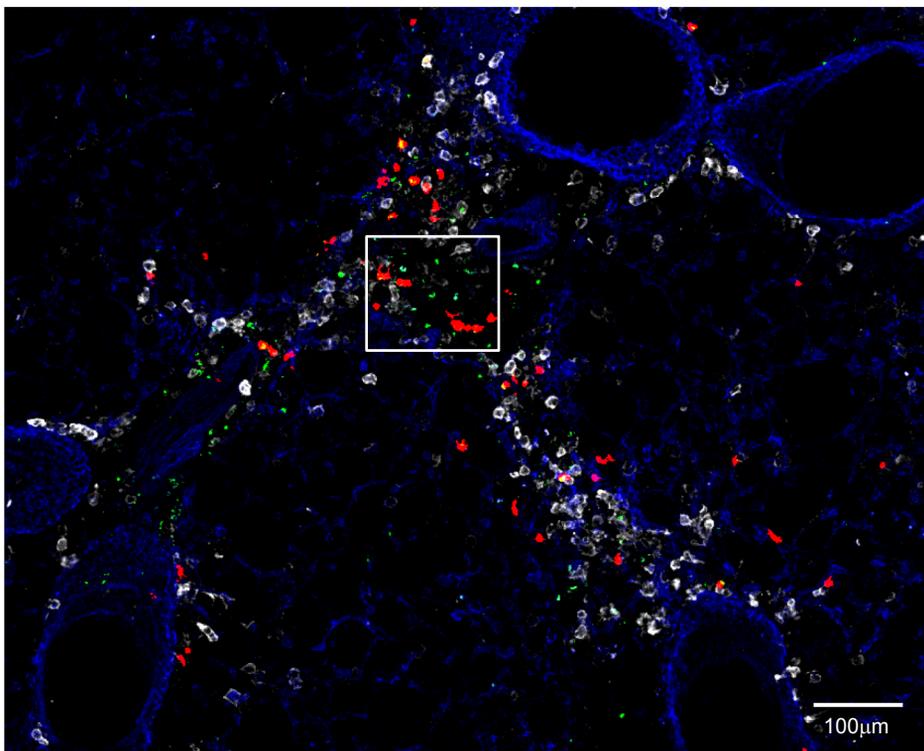
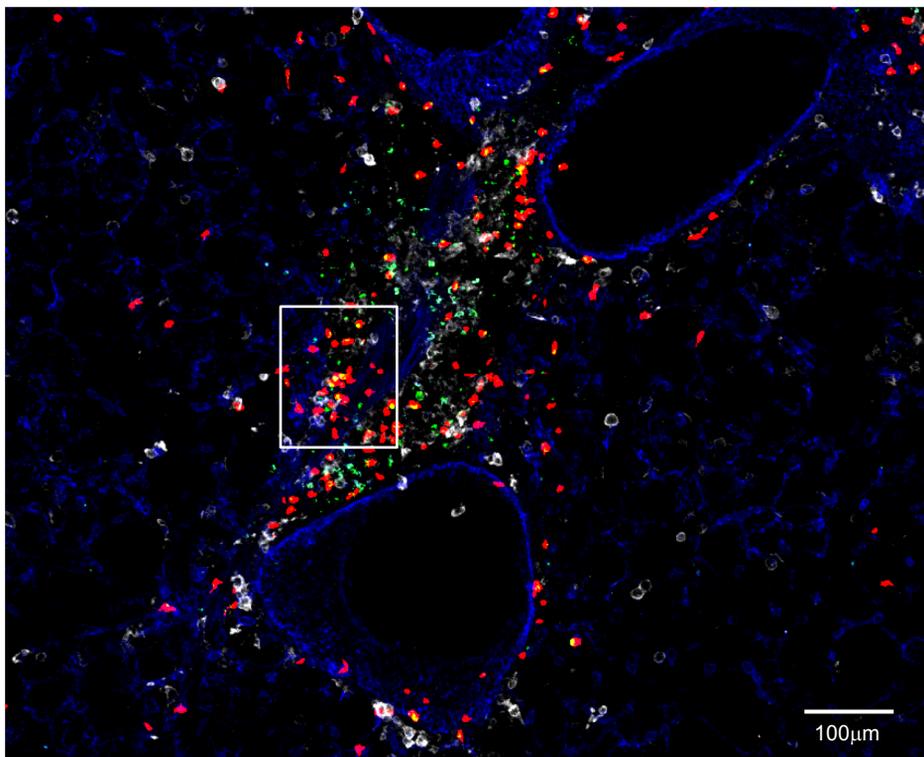
Supplementary Figure S3. Identification by CyTOF of cell populations in the lungs of OVA-challenged mice. (Top) IL-17 staining of cells within cell populations identified as Th17 cells (1), other CD4⁺ T cells (2), CD8⁺ T cells (3), TCR $\gamma\delta$ ⁺ T cells (4), neutrophils (5), NK T cells (6), CD103⁺ DCs (7), CD11b⁺ DCs (8), interstitial macrophages (9), alveolar macrophages (10) and B cells (11). Also shown are staining for molecules that contribute to the identification of these cell populations.

Supplementary Figure S4



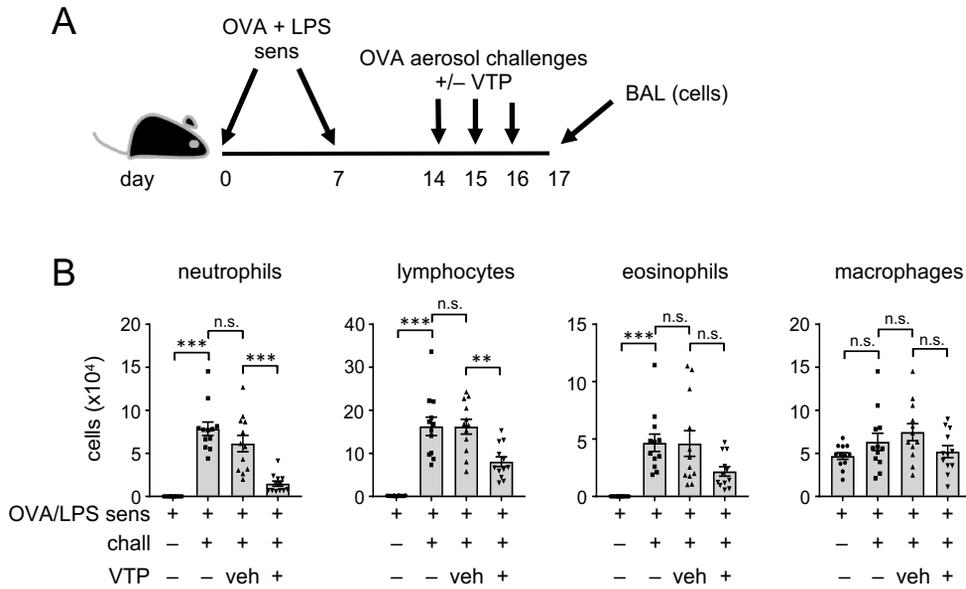
Supplementary Figure S4. Gating strategy for IL-17-producing cells in the lung. Shown are gating strategies to identify IL-17⁺CD3⁺ T cells (P1), IL-17⁺ non-T cells (P2), IL-17⁺CD4⁺TCR $\alpha\beta$ ⁺ T cells (P3) IL-17⁺CD4⁻TCR $\gamma\delta$ ⁺ T cells (P4), interstitial macrophages (P5) and alveolar macrophages (P6).

Supplementary Figure S5



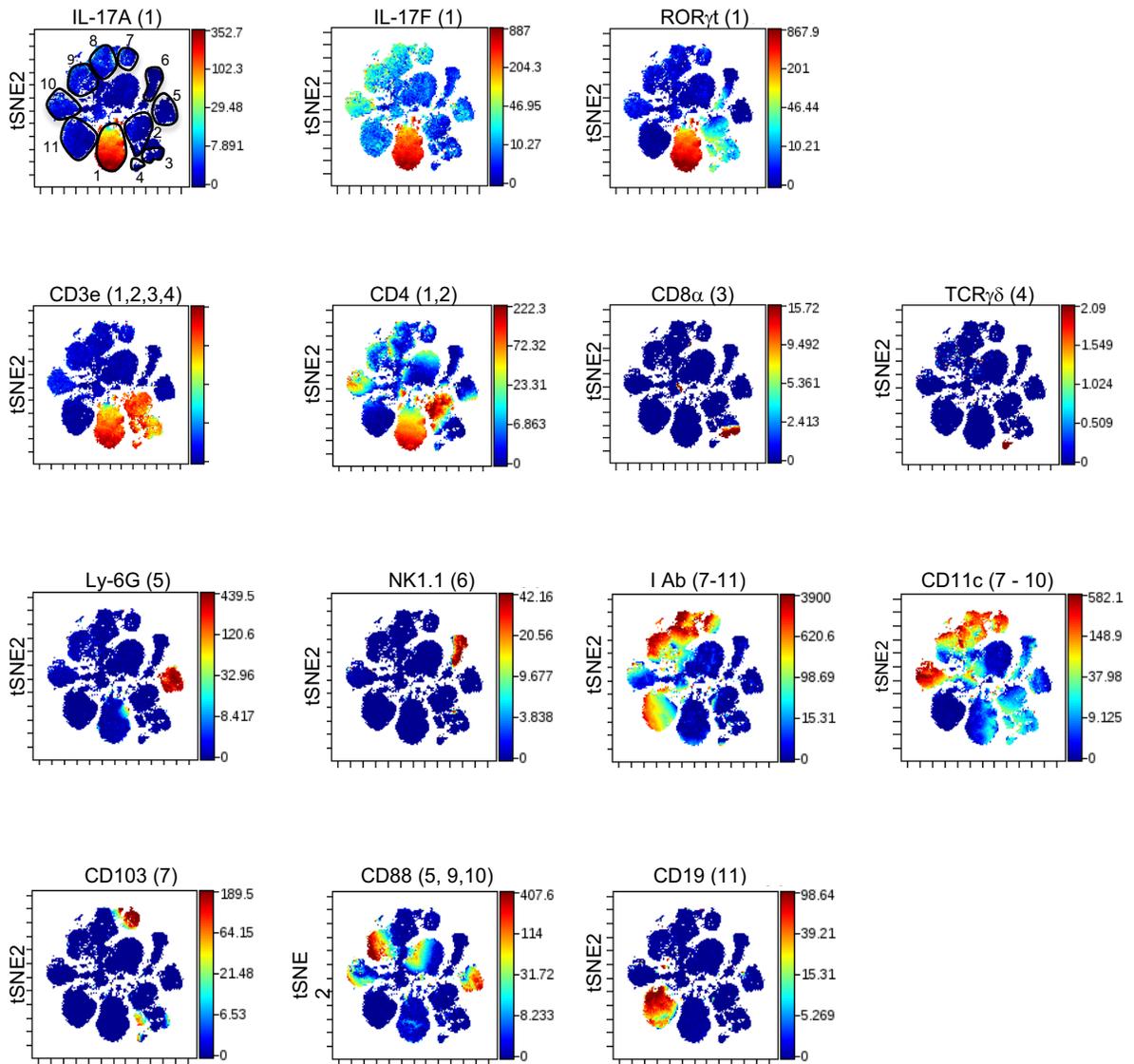
Supplementary Figure S5. Confocal image of PCLS showing localization of Th17 cells. Shown are Th17 fate mapping cells (red), IL-17 (green), IL-17-expressing Th17 cells (yellow), CD11c⁺ APC's (white) and epithelial cells (blue). Images are from vehicle-treated mice (top), and VTP-treated mice (bottom). White rectangles represent areas shown at higher power in Figure 4D. Data shown are from a single experiment, representative of two.

Supplementary Figure S6



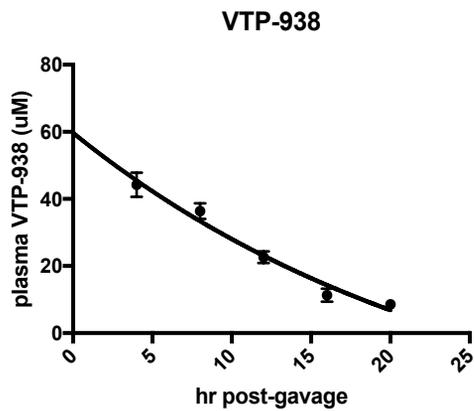
Supplementary Figure S6. Effect of VTP given prior to allergen challenge on three consecutive days. (A) Timeline for OVA/LPS-mediated sensitization of mice given VTP or the vehicle (veh) during the challenge phase (3 consecutive days of OVA aerosol). **(B)** Numbers of cells corresponding to the indicated leukocyte subsets in BAL at 24 h post-challenge. *N*=12 mice/group; data are combined from two experiments. Kruskal-Wallis 1-way ANOVA with Dunn's multiple comparison test. n.s.; not significant. **P*<0.05, ***P*<0.01, ****P*<0.001.

Supplementary Figure S7



Supplementary Figure S7. Identification by CyTOF of cell populations in the lungs of HDE-challenged mice. (Top) IL-17A, IL-17F and ROR γ t staining of cells of cell populations identified as Th17 cells (1), other CD4⁺ T cells (2), CD8⁺ T cells (3), TCR γ δ ⁺ T cells (4), neutrophils (5), NK T cells (6), CD103⁺ DCs (7), CD11b⁺ DCs (8), interstitial macrophages (9), alveolar macrophages (10) and B cells (11). Also shown are staining for molecules that contribute to the identification of these cell populations.

Supplementary Figure S8



Supplementary Figure S8. Plasma concentrations of VTP-938. Mice were gavaged with 30 mg/kg VTP-938, and blood was collected at the indicated times post-gavage.

Metal label	target	clone	Manufacturer	Catalog #
141Pr	Ly-6G	1A8	Fluidigm	3141008B
142Nd	CD11c	N418	Fluidigm	3142003B
143Nd	Biotin	1D4-C5	Fluidigm	3143008B
144Nd	FITC	FIT-22	Fluidigm	3144006B
145Nd	PE	PE001	Fluidigm	3145006B
148Nd	CD11b (Mac-1)	M1/70	Fluidigm	3148003B
149Sm	CD19	6D5	Fluidigm	3149002B
151Eu	CD25 (IL-2R)	3C7	Fluidigm	3151007B
152Sm	CD3e	145-2C11	Fluidigm	3152004B
153Eu	CD103	2E7	Fluidigm	3999999-1
154Sm	Siglec-F	E50-2440	Fluidigm	3999999-1
155Gd	CD326 (EpCAM)	G8.8	Fluidigm	3999999-1
156Gd	CD88	20/70	Fluidigm	3999999-1
159Tb	RORr(t)	B2D	Fluidigm	3159019B
160Gd	CD62L (L-selectin)	MEL-14	Fluidigm	3160008B
163Dy	IL-17F	D3M4D	Harvard*	N/A
165Ho	TCRgd	GL3	Harvard*	N/A
168Er	CD8a	53-6.7	Fluidigm	3168003B
170Er	CD161 (NK1.1)	PK136	Fluidigm	3170002B
171Yb	CD44	IM7	Fluidigm	3171003B
172Yb	CD4	RM4-5	Fluidigm	3172003B
174Yb	IL-17A	TC11-18H10.1	DVS Sciences	3174002B
175Lu	IAb	AF6-120.1	Fluidigm	3999999-1
176Yb	APC	APC003	Fluidigm	3176007B
89Y	CD45	30-F11	Fluidigm	3089005B
Non-metal label	target	clone	Manufacturer	
Biotin	CD90.2/Thy-1.2	53-2.1	eBioscience	13-0902-82
APC	CD196 (CCR6)	29-2L17	R&D	FAB590A
FITC	CD31 (PECAM-1)	390	BD Pharmingen	553372
PE	F4/80	BM8	eBioscience	12-4801-82

* Harvard Medical Area CyTOF Antibody Resource and Core, Boston, MA

Supplementary Table S1. List of antibodies used for CyTOF experiments.